

Introduction

simplifying research

Downstream of B-Cell Receptor (BCR)/CD40 signalling, the PI3K-AKTmTOR axis promotes cell survival and proliferation via negative regulation of FOXO proteins. FOXOs are transcription factors belonging to the Forkhead Box (FOX) superfamily typically described as canonical 'tumour suppressors' due to their roles in pro-apoptotic and anti-proliferative responses

We have previously demonstrated that synergistic inhibition of BCR ligation using AZD8055 (dual mTOR inhibitor) and Ibrutinib (BTKi) leads to CLL cell death coinciding with increased FOXO1 activity (Cosimo et al., 2019), thereby demonstrating FOXO1's tumour suppressive capability. However, unlike other B-cell malignancies, wider FOXO family function in CLL is poorly described (Lees *et al.,* 2023). Here we describe a potential role for FOXO4 in CLL proliferation and survival, focusing on an shRNAbased approach to investigate the characteristics of FOXO4, as well as exploring FOXO4 expression and localisation in distinct CLL primary and cell line models.



Normal conditions (NDC)



Objectives

To provide insight into novel FOXO4-mediated regulation of intracellular mechanisms following in vitro stimulation, drug treatment and shRNA-mediated FOXO4 depletion.



NTL-CD40L Co-culture

- NTL-CD40L in vitro co-culture, $F(ab')_2 \bigcirc \bigcirc \bigcirc$ stimulation
- Western blotting (WB)
- *Ex vivo* samples, 1hr, 48hr treatments & cellular fractionation
- **RNA Sequencing**
- 5 patient samples, 24hr treatments 2. RT-qPCR
- FOXO4, BCL2L11, SESN3, GADD45A expression (24 hr), cell line & patient samples
- 3. shRNA-mediated FOXO4 knockdown (KD) Cell lines & patient samples
- 4. Flow cytometry
 - CTV, Annexin/7-AAD, i/c pH2AX



NTL-CD40L primary CLL co-culture

Drug treatment

FOXO4 is Required for the Promotion of CLL Growth and Survival Jamie Lees¹, Jodie Hay¹, Alison McCaig², Alison Michie¹

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Figure 1. (A) (left) WB depicting the basal expression levels of FOXO4 in HG3 and MEC1 cell lines, which possess cytogenetic alterations associated with either favourable or poor prognosis respectively. (right) displays quantified intensity levels of FOXO4 expression between the two cell lines (n=3). (B) Protein lysates were prepared from *ex vivo* PB CLL patient samples and were processed via WB to detect FOXO4 expression from a wider cohort of patients (n=18) vs. healthy B-cell donors (n=5). The blots show differential FOXO4 expression, with a significant increase of FOXO4 expression seen within a Binet Stage A subgroup of CLL patients (A. n=4, B. n=7, C. n=7)

Stimulation promotes increased FOXO4 expression and further nuclear localisation <u>Stim US Stim</u>



Figure 2. (A) A representative WB showing how CD40L stimulation of primary patient samples increase FOXO4 expression in vitro. (B) Subcellular fractionation of primary patient samples demonstrates an abundance of nuclear FOXO4 that increases following either CD40L or BCR-stimulating F(ab')₂ fragment). (C) Subcellular fractionation of HG3 cells supports a persistent nuclear abundance of FOXO4. Lamin A/C and GAPDH are used as loading controls to detect sufficient fractionation of nuclear and cytoplasmic fractions respectively.





Results

FOXO4 expression is tightly controlled during CLL cell proliferation





Figure 3. (A) (left) Volcano plot representing differentially expressed genes in an RNA-Seq dataset profiling 5 proliferating patient samples (+CD40L) in response to AZD8055 treatment. (right) A heatmap describing regulation changes within members of the FOXO signalling pathway with *FOXO4* being the most significantly upregulated. (B) Changes in FOXO4 expression in response to CD40L stimulation, AZD8055, ibrutinib or combination treatment (n=5, 24 hr). (C) Validation in MEC1 cells mimics FOXO expression increases seen in the RNA-Seq dataset +AZD8055 (n=3). (D) RT-qPCR reveals a reduction in *FOXO4* expression (+CD40L) in primary cells that significantly increases following 10 days in in *vitro* co-culture (n=6).

FOXO4 plays a crucial role in inhibiting CLL chemosensitivity



CLL cells are more susceptible to DNA damage following FOXO4 KD



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Conclusions

HG3

Figure 4. (A) Scatter plots depicting increased cell death in MEC1 (top) and primary CLL (bottom) cells with FOXO4 KD following AZD8055-Ibrutinib combination treatments. (B) Significant increases of Annexin⁺/7AAD⁺ MEC1 and HG3 *FOXO4*-KD cell populations following drug treatment (n=4 MEC1, n=3 HG3). (C) qPCR and ■ Untreated AZD8055 +Ibrutinib WB analysis of the proapoptotic FOXO target gene *BCL2L11* and its protein product BIM, supporting an increase in apoptotic cellular environment AZD8055 + Ibrutinib AZD8055-Ibrutinib treatment (MEC1 cells, n=4 qPCR, n=3 WB).

- High nuclear abundance of FOXO4 and localisation in stimulated patient CLL cells points toward basally 'active' FOXO4 in vitro, compared with FOXO1 in CLL being 'inactive' (Cosimo *et al.*, 2019).
- Significant changes in FOXO4 expression in response to AZD8055-Ibrutinib treatments as well as throughout CD40L co-culture demonstrates how CLL survival and proliferation is partnered with discrete FOXO4 regulation
- Increases in CLL cell chemosensitivity following shRNA-mediated FOXO4 depletion suggests a novel requirement for FOXO4 abundance to promote CLL proliferation and survival.
- Varied levels of FOXO4 in distinct CLL subgroups demonstrates a potential link between FOXO expression and disease prognosis



References

Cosimo et al. (2019): AKT/mTORC2 Inhibition Activates FOXO1 function in CLI Cells Reducing B-Cell Receptor-Mediated Survival. Clin Cancer Res. doi 10.1158/1078-0432.CCR-18-2036

Lees et al. (2023): The Discrete Roles of Individual FOXO Transcription Factor Family Members in B-Cell Malignancies. Front. Immunol. doi: 10.3389/fimmu.2023.1179101

